

There is 3 major staining techniques

① **Histochemistry**: staining by reactions between tissue components & chemicals

includes (the most common)

* **H&E** (Hematoxylin & Eosin)

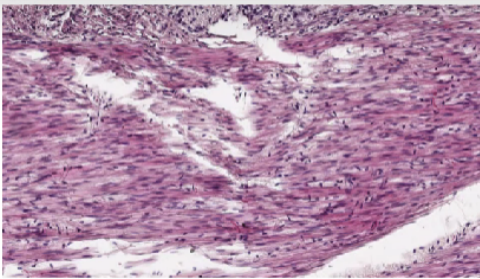
Stains nucleus **blue** (cause Hematoxylin is base and the nucleus contains Acids (DNA))

Stains cytoplasm **pink** (cause Eosin is Acidic and the cytoplasm contains many Basic proteins)

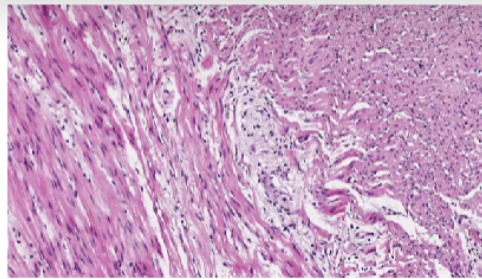
note: Eosin varies (pink to orange including red) / Hematoxylin (blue to purple)

According to: ① How long the specimen has been preserved
② How long it stayed in the stain solution

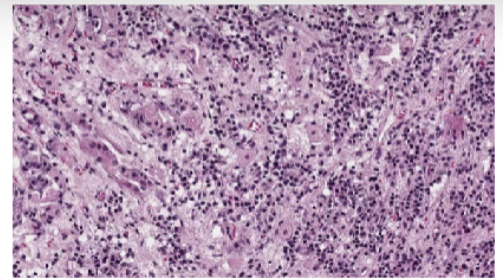
Histological staining: hematoxylin & eosin



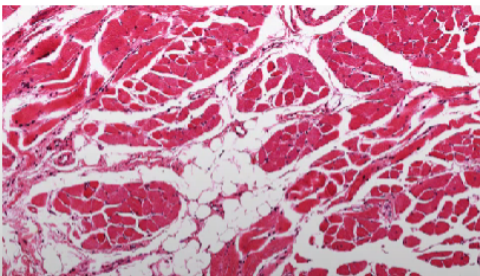
Mayer's hematoxylin (5 min)
+ Eosin aqueous (3 min)



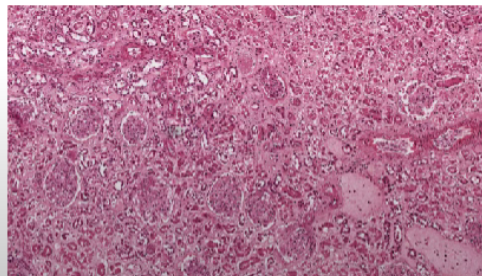
Mayer's hematoxylin (8 min)
+ Eosin aqueous (3 min)



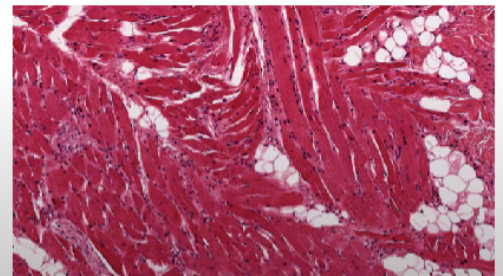
Mayer's hematoxylin (10 min)
+ Eosin aqueous (5 min)



Mayer's hematoxylin (5 min)
+ Eosin alcoholic solution (30 sec)

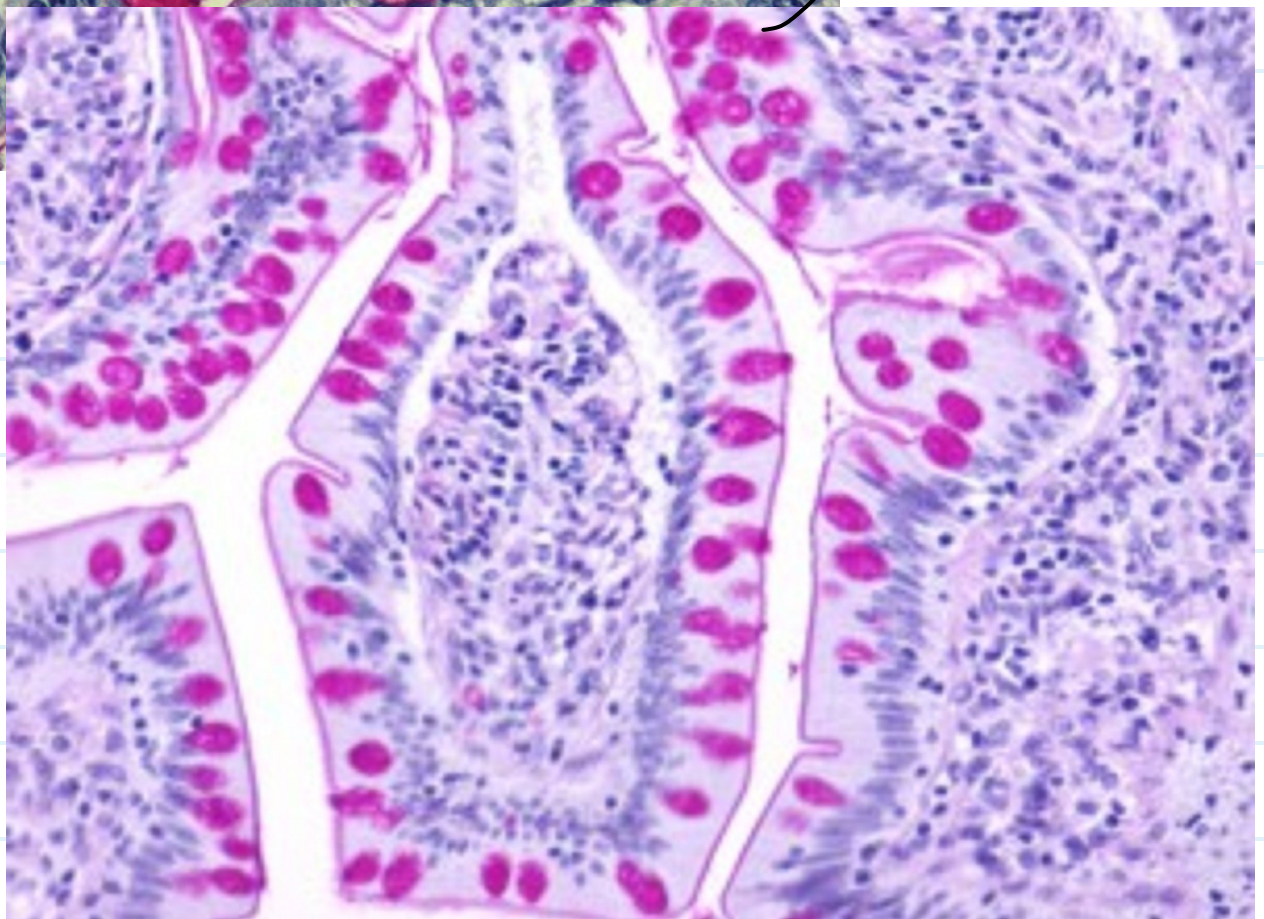
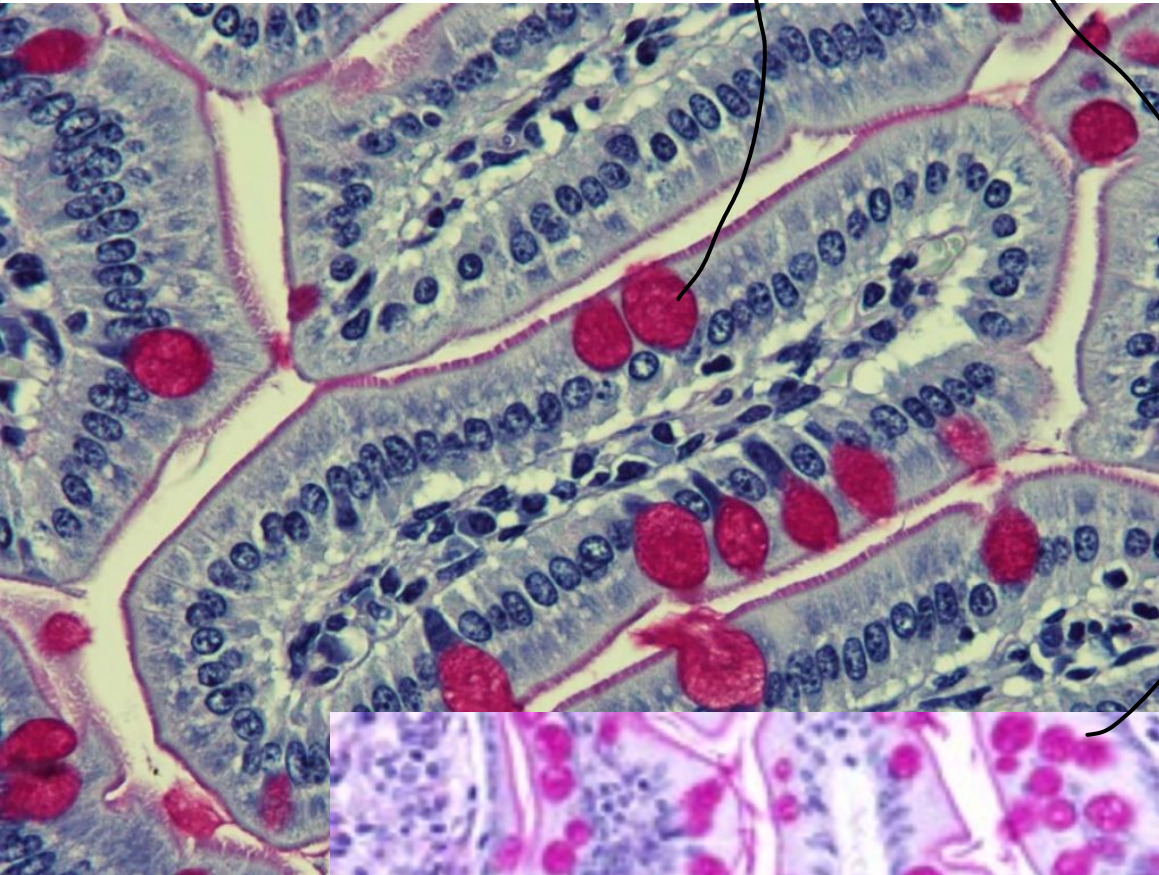


Mayer's hematoxylin (8 min)
+ Eosin alcoholic solution (60 sec)

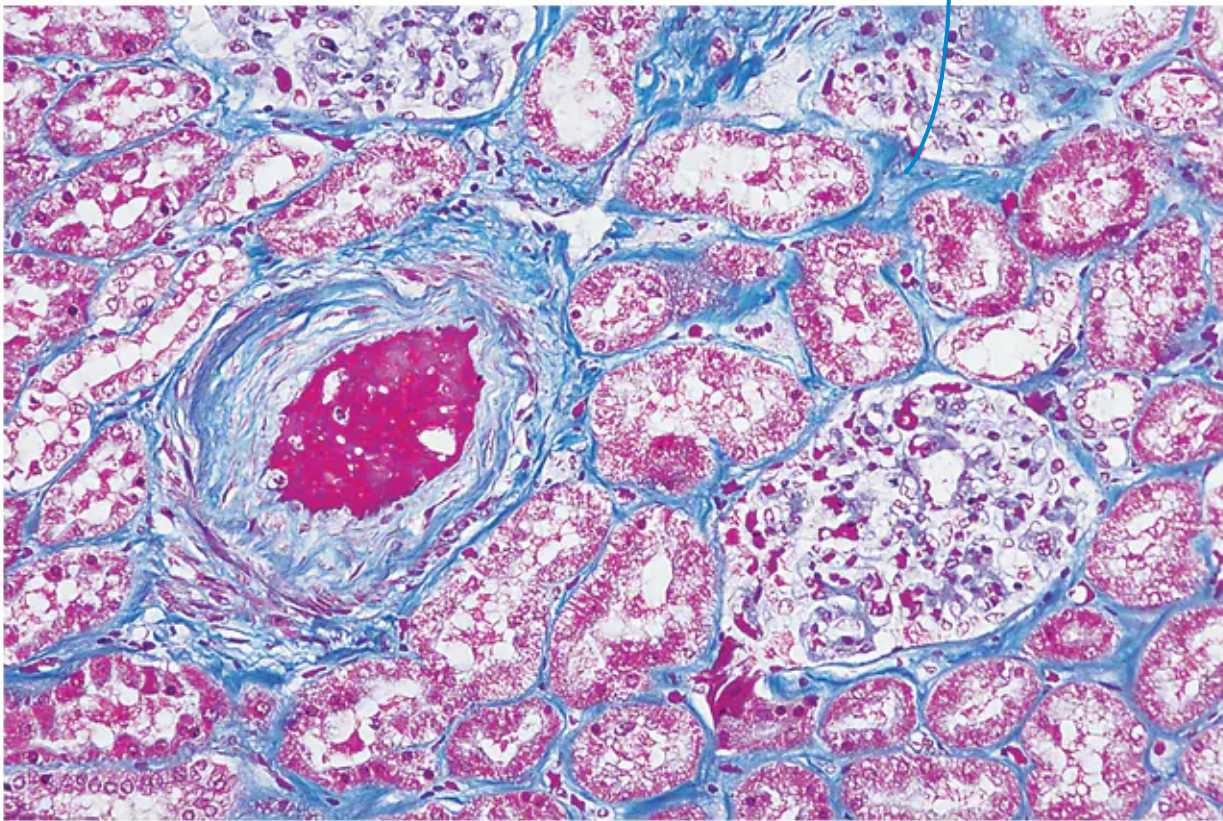
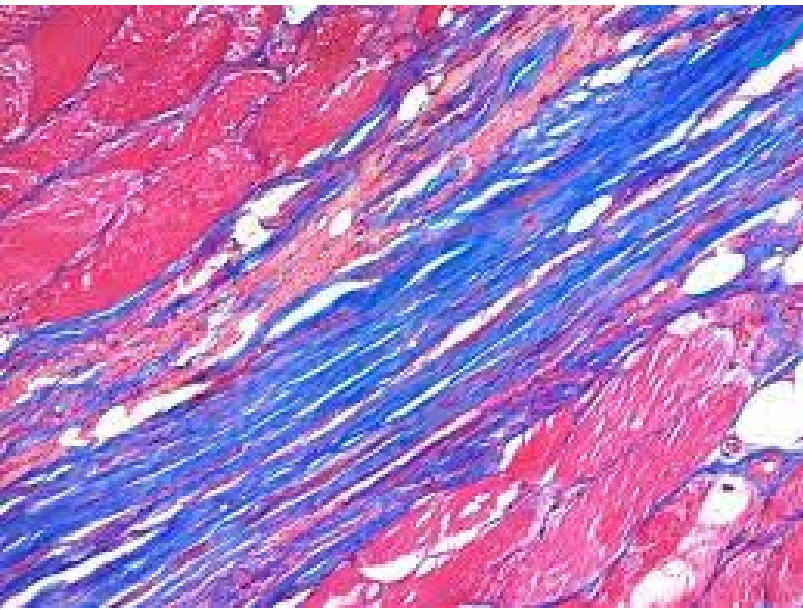


Mayer's hematoxylin (8 min)
+ Eosin alcoholic solution (90 sec)

PAS (Periodic Acid Schiff) : Usually reacts with
. Carbohydrates to visualize it
and it appears Red or Magenta



Trichrome: combination of dyes often used
3 colors to visualize collagen (which appear blue)



2 Immunohistochemistry:

using specific interactions between Antigens & Antibodies

* used to stain certain proteins

How it is done:

Directly: ① An Antibody with chromogen (*color generating Antigen*)

is attached to the protein after recognizing it

② certain chemical is applied to react with chromogen

to turn the tagged region $\xrightarrow{\text{to}}$ Brown color

In Directly: here an additional step happens:

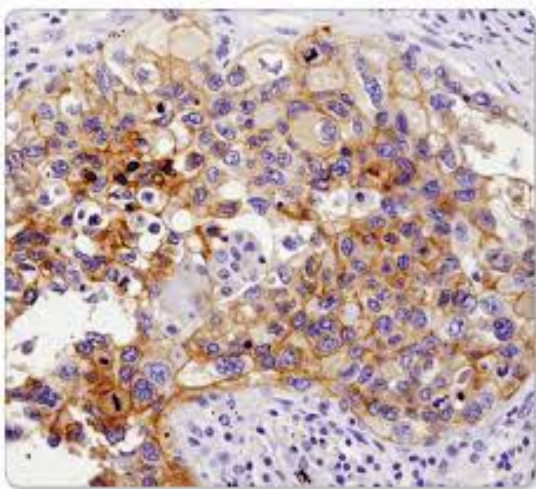
Rather than using the same Antibody to recognize the protein

& Hold the chromogen, primary Antibody recognize

and a secondary Antibody with chromogen attach

to the primary Antibody.

Then the same chemical is applied



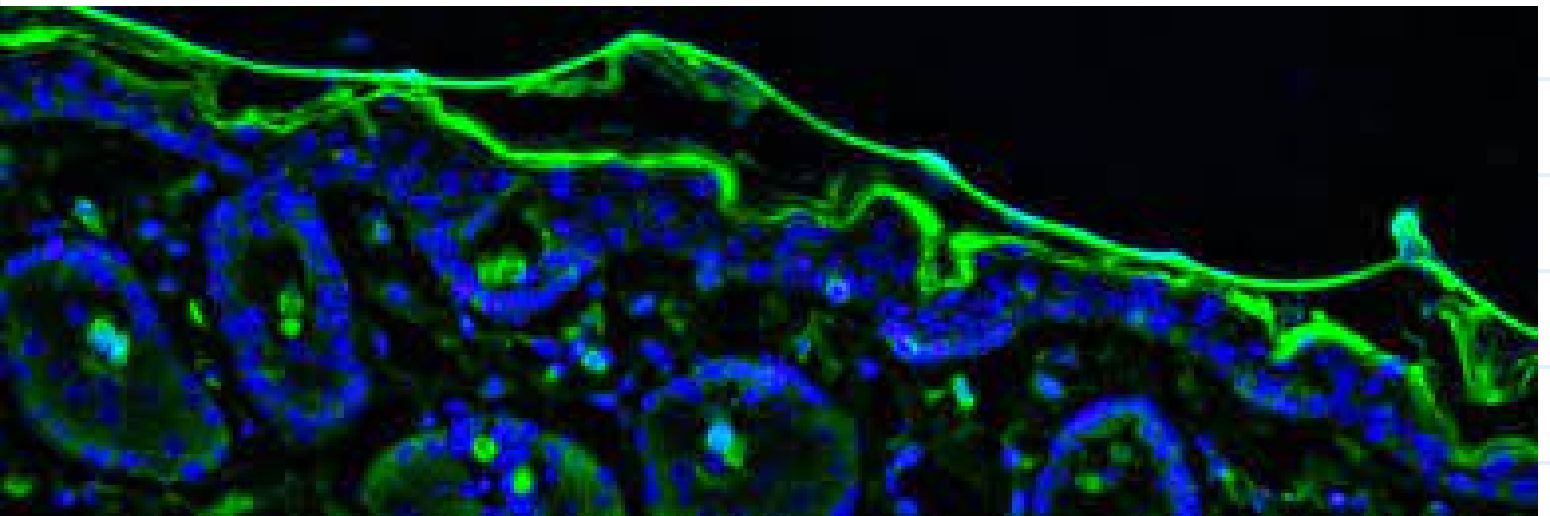
3] Immunofluorescence:

Just like the previous one with (Fluorescent Tag) instead of chromogen.

Also the Fluorescent Tag is excited by UV light or Laser not a chemical substance

This excitation causes the Fluorescent Tag to emit specific frequency in the light spectrum

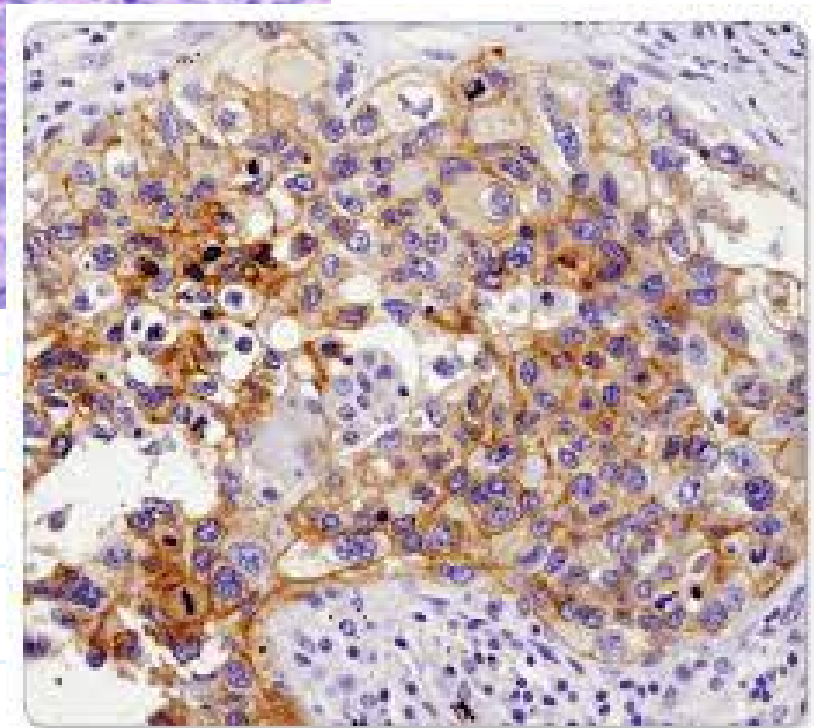
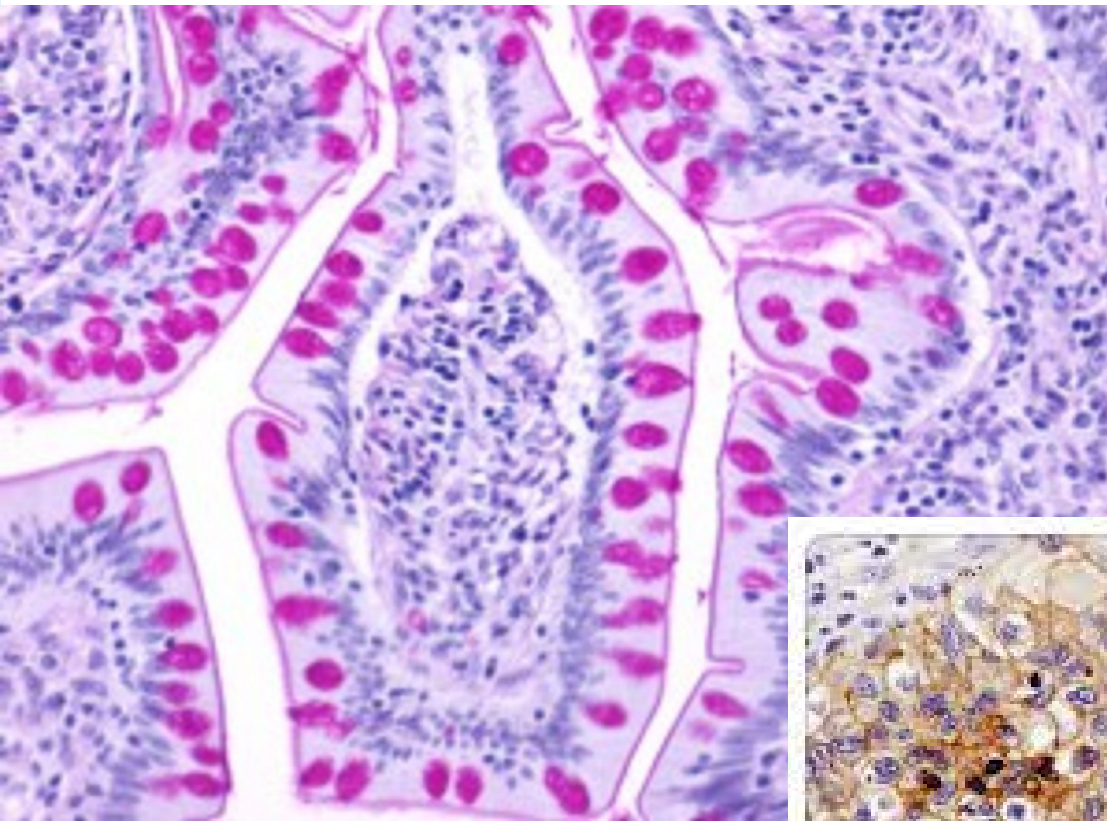
* Common colors are Green & Blue on blackground
(microtubules) (nuclear proteins)



Light microscopes

1 Bright-field :

- *white background
- *used to observe tissue stained with histochemistry (H&E, PAS, Trichrome)
- *also it is used to observe tissue stained with Immunohistochemistry

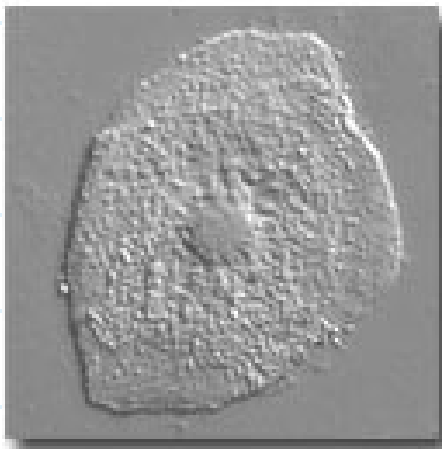


2 Phase-contrast:

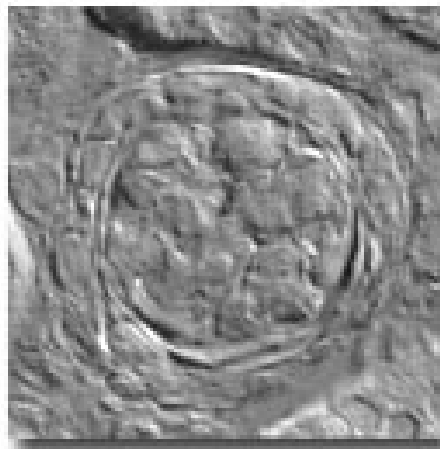
* Gray background

*used to observe (non-stained) tissues
so it can be living tissue or cultured cells

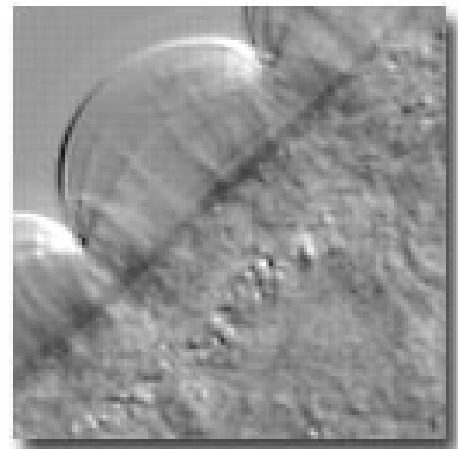
Transparent Specimens in Phase Contrast and DIC



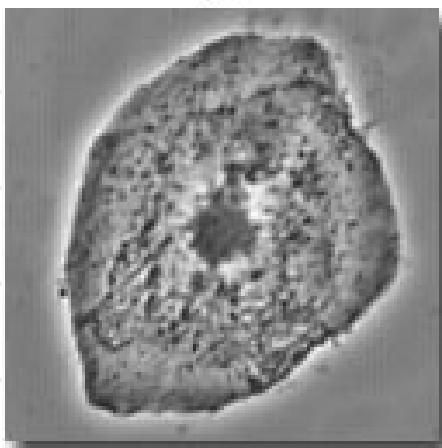
(a)



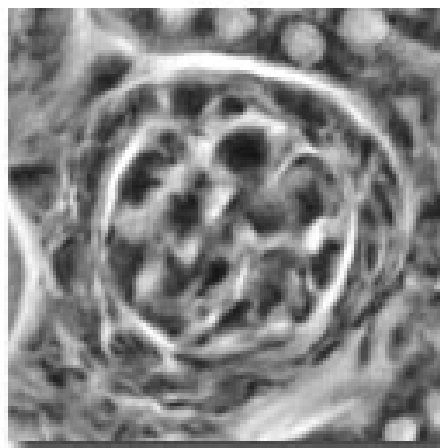
(c)



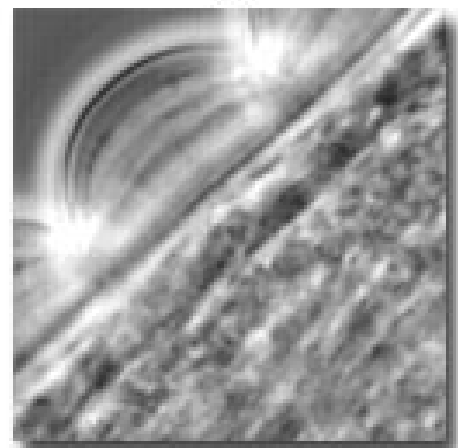
(e)



(b)



(d)



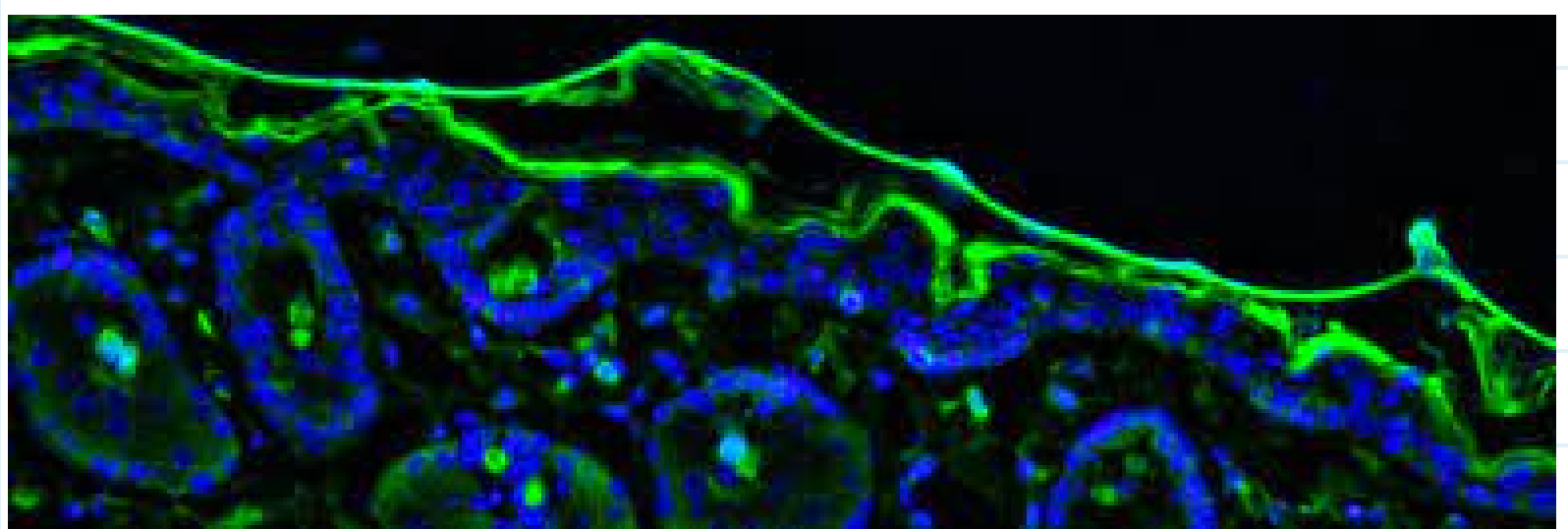
(f)

Figure 1

Fluorescence microscopy :

* Black background

*used to visualize Immunofluorescence stained tissues



Electron microscopes:

1 TEM:

*Greatest magnification & resolution

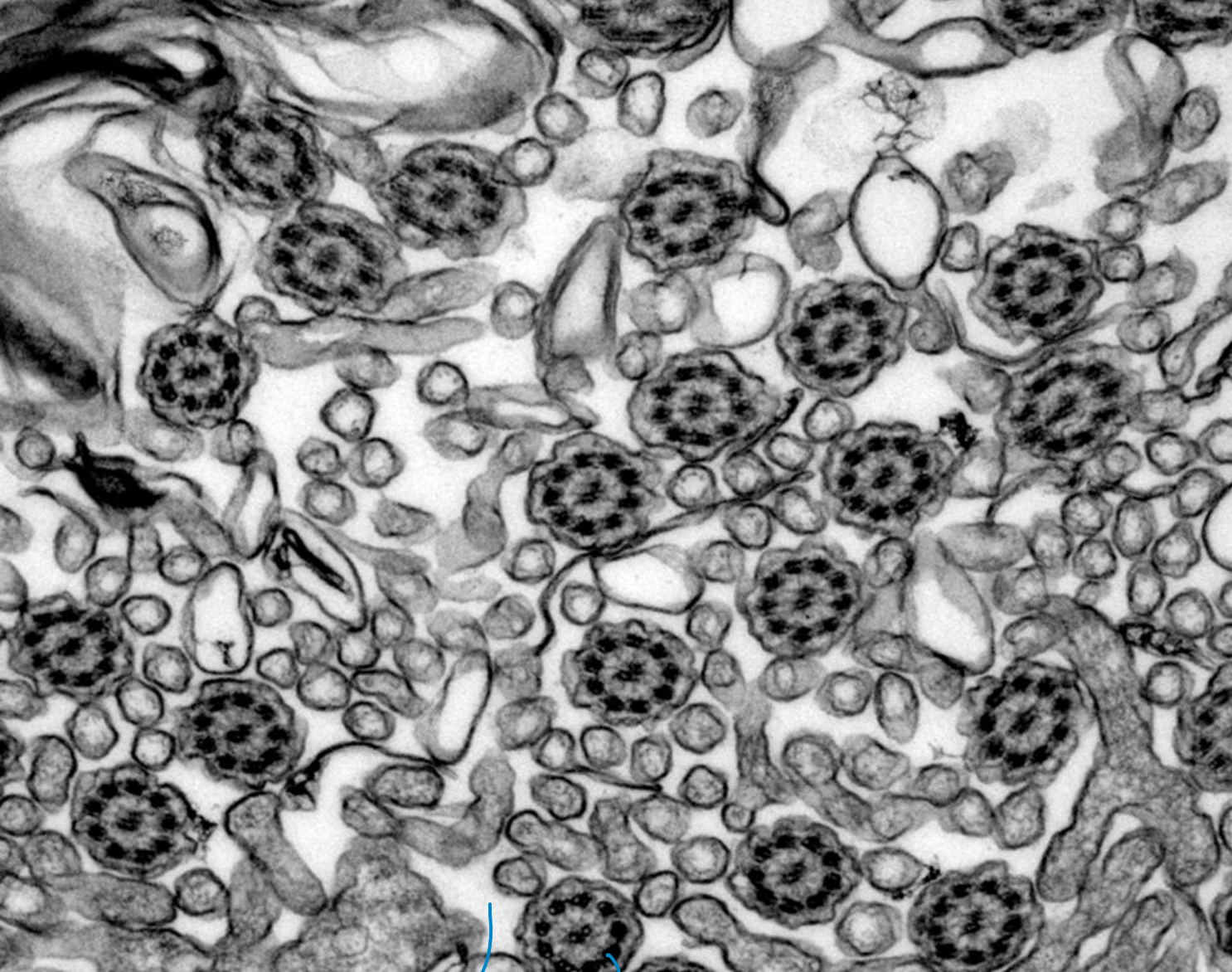
*electron beam passes through the tissue (the specimen)

*Images colors are shades of Gray :

▶ the Darker Areas (dense electron area)
not enough electrons were able to pass
so cells or tissues are concentrated

▶ the lighter Areas (electron light area)
a lot of electrons were able to pass

so fewer cells or tissues are concentrated there
(so it is most likely to be fluid)



Light Areas

Dark Areas

2 SEM:

*electron beam does not pass through the tissue (the specimen)

*The surface of cells & tissues are coated with heavy metals (usually gold) which reflect electrons

* Generate 3D images

they are originally colorless but they can be colored using computer programs

